

PerkinElmer Operetta CLS



Overview



Service contact (administrators only): "EVO Portal", Perkin Elmer website.

Incubation: Heating is available (37-42 °C; system temperature of about 25 °C without heating activated). CO₂ controller available (1-10 %). No humidity control: Use closed samples in excessive imaging buffer, if necessary.

Objectives: 5x, 20x, 40x (all air).

Light/optics: 4x LED light source (can activate 1 source simultaneously), 5x emission filter (UV / green /yellow / red / far-red), Spinning disc optics. 1x top-down-LED (740 nm, for morphology, using transparent-top samples).

Camera: 4.7 Megapixel sCMOS sensor.

Light path/wavelengths

Excitation windows available:

- **370 nm** (355-385 nm)
- **475 nm** (460-490 nm)
- **550 nm** (530-560 nm)
- **630 nm** (615-645 nm)

Detection windows available:

- **430-500 nm** (blue)
- **500-550 nm** (green)
- **515-580 nm** (yellow)
- **570-650 nm** (red)
- **665-760 nm** (far-red)



What about my specific excitation line?

For fluorophores with an excitation peak at for example 510, use the closest available light source: 475 nm
See <https://searchlight.semrock.com/> for excitation/emission spectra of many fluorescent dyes and proteins.

When testing a new assay, make sure to include enough controls to verify that no crosstalk appears.

Plates

- Manufacturer comments on plate usage: <https://www.perkinelmer.com/uk/lab-products-and-services/application-support-knowledgebase/microplates/microplates-knowledge-base.html>
- Machine takes "all" available cover-slip bottomed plates.
- But, do not use Techno Plastic Products (yellow stripe); which have plate-vents and cannot be supported by the holder.
- may have a glass or plastic "coverslip" bottom
- for good cell-attachment, can coat the bottom with poly-L-lysine
- When correctly entered, objectives will never touch the plate bottom (if still; plate will be pushed upwards).



Skirt height

Difference between plate-edge height, and imaging surface height
Important value to enter into machine: 0-1.5 mm range allowed.
Can measure the height of unknown plates using the Operetta with the 20x objective.

Edge effects

Prefer not to use the outer rows and outer columns of any given plate.
This because of "edge effects" (thermal, evaporation, different near the plate edge).
Fill those wells with only buffer, for example, and do not use for imaging.

If the edge wells are needed for samples too, make sure to fill the inter-well space, which reduces edge effects in case of 96-well plates (see http://resources.perkinelmer.com/lab-solutions/resources/docs/tch_perform_successful_long_term_live_cell_imaging_in_high-content_analysis_system_013900_01.pdf).

Usage

Machine start

- Depress button (green) on the left side of the machine
- Wait a few minutes until the software displays "device is ready".
- The machine is now ready to use

Around the sample area

- If the table is not powered, eject/re-insert the plate holder (software). Do not manually move table.
- Plate/sample holder is mounted on steel tracks: DO NOT CLEAN/TOUCH (if dirty, call service)
- Plate/sample holder contains an internal "penta-pattern" of dots, used for high-accuracy positioning of the sample.
- Red dot on objectives needs to match the red dot on the microscope. Objectives can be manually lifted out of the system.
- With open lid, the machine counts as a class 3B laser. Close the door firmly.
- Emergency release hatch button (lid open) is on the lower right of the machine.

Plate loading

- Start Harmony software
- Log-in with user account: personal account, with personal settings
- User accounts can only delete own data. Administrators can delete all.
- Choose plate. Nomenclature = number of wells, manufacturer, type/name of plate
- Choose objective. Start at eg. 20x / 0.4 NA
- Choose operational mode (opt-mode). Confocal = spinning disc mode, with pinholes, 5% transmission of light, both ways. Epi = non-confocal, 100% transmission of light, both ways.
- Binning: (makes image size <4.7 Megapixel) signal x4, noise x2, resolution significantly reduced.
- Live preview: eg. Show current image during stack acquisition. off = evaluate manually after setting up acquisition.
- Click "new" new experiment.
- Set up individual channels for your fluorophores.



Arrowheads

Located under menus always reveal more detailed settings.

- Plate layout (right-most). Select wells to measure, click "SELECT". Select FOV, number, position.
- Select well-to-test.
- Clean plate-bottom with wipe (KimTech or similar) and 70% Ethanol before use.
- Load plate with or without lid (place within protrusions).
- or: Load slides in holder.
- or: Load slide holder into Operetta.

Setting up for imaging

- Set up the desired channels, select a well, and take a snapshot. Different z-heights may need to be trialled.
- Evaluate brightness; auto-scale and auto-contrast are always applied.
- No range-indicator (over/under exposure) is available, but a histogram for each channel is displayed.
- Histogram: if not visible, can be enabled using the arrow under "Channels" to the right of the image.
- Scaling: Normal mode: Black-to-Colour. Enhanced mode: Black-to-Colour-to-White.
- Right-click on the sample image, "show intensity". Ctrl + Click to add ROIs.
- Find focus height: Z-stack! "test" is used here, and only applies to the well that was highlighted by clicking, on the right.
- Start at negative values. Go up to positive. See in "TEST IMAGES" (right of screen)
- Pick the right focus height for each channel individually.
- Next, to be certain, do focus finding in another cell. Possible plate-tilting may be revealed, and needs inspection.



Z-stacks yes/no?

It is possible to disable "use z-stack in test", to test with only single-plane images
If you want no stack acquisition upon running the actual experiment, simply select "0 planes"

"Selecting" wells and highlighting wells

Clicking on wells highlights them, and points at wells to be used in test images
Selecting wells (highlighting, then clicking "select") determines which wells to use in the experiment.

- Overview picture: in the "well layout" section, select all interesting wells; right click "overview;" preview field: observe the stitched image afterwards.
- The overview picture is automatically downscaled to 200 MB in size; full-resolution stitching is not supported.
- Double-check overlap: Pull up the contrast to an extreme value (visually; right-click in the image)
- Determine whether flat-field correction will be necessary.
- "online-job" = perform image analysis during acquisition
- You can add layouts (labels) to the output data. This is an additional table.
- Re-use a plate layout (saved, from eg. editor or previous experiment); "Assay" in "define layout" (right-most).



Use overview image to pick interesting regions

Using "Test Images", and marking regions, right-click and "use as background for well" for a visual aid for selecting further regions.

Run the experiment

- Evaluate that the correct assay is loaded.
- Name the plate and add keywords.
- **Start!** And observe live-preview images during acquisition (when enabled)
- Select the whole plate, right-click, organize images in "realistic" (corresponding to the plate) or "packed" (best for display) layouts



Rule of thumb for estimated dataset size

The Operetta CLS is only displaying the total size of a dataset if **not** enough disk space is available. If you want to know the estimated dataset size before you start an experiment, use the following rule of thumb:

Number of wells x fields x channels x z-planes x timepoints x 8 MB = approximate dataset size in MB



Flat-field correction

The software can do flat-field correction based on acquired images, provided that foreground/background and areas without cells are present. This also works in wells filled with tissue, or more-than-confluent cell cultures.

- When done: eject the sample (settings - Operetta CLS - eject) and remove plate/slide holder. Afterwards "eject" once more to close the system again.

Exporting data

- In the "settings" menu up top, data management can be used to export data
- Export your data as an "archive" for later re-analysis using Harmony.
- Export your data as "measurement + associated data" to save results as TIFF files, alongside already-performed measurement outcomes, and re-analysis eg. using CellProfiler. Be aware that the exported raw images are **not** flat-field corrected, so it might be necessary to correct the images before the analysis.
- Connect to the CAi shared network drive to store the data, or bring your own data-carrier (USB).
- After the session, delete your data from the Operetta computer.



Saved parameters

During an experiment, you define and save the following objects:

- *Experiment* (light, stacks, channels, etc)
- *Assay layout* (cells, treatments, rows/column names)
- *Measurement* (image data)
- *Analysis sequence* (processing steps, calculations, values of interest)
- *Evaluation* (grouping, graphs/charts, heatmaps, well-display, statistics, data points)

Image processing and analysis



Loose info pieces:

Z-steps always > 500 nm

Fluorescent staining strategy:

To facilitate auto-detection in subsequent image analysis steps:

- a)** stain cells with target marker
- b)** stain cytosol separately (eg. CellMask Blue)
- c)** stain nuclei separately

Computer attached to the microscope:

DO NOT SWITCH OFF!

DO NOT UPDATE WINDOWS!

The PC is running a live image data and image analysis server.